

Streptococcus mutans in denture stomatitis patients under antifungal therapy

Streptococcus mutans em pacientes com estomatite protética submetidos à terapia antifúngica

Abstract

Purpose: To assess the number of *Streptococcus mutans* in saliva of patients with denture stomatitis before and after antifungal therapy.

Methods: After examining 93 patients, 47 were selected for fungal test. Then, from this sample, thirty patients were selected: 15 with positive and 15 with negative diagnosis for candidiasis that were evaluated for *S. mutans* counting, salivary flow and buffer capacity evaluation. Oral hygiene and prosthesis hygiene, period using prosthesis, lesion type and salivary data were related with clinical laboratorial characteristics of the patients with *Candida*.

Results: The most frequent lesions were type I (43.5%) and II (53.5%). The amount of *S. mutans* was six times higher in patients with candidiasis and it was associated with low salivary flow and poor oral hygiene. After therapy, a reduction of *S. mutans* was verified particularly in patients with normal salivary flow. The values ranged from 0.01 to 3.88×10^4 cfu/mL.

Conclusion: The data suggest that *Streptococcus* spp collaborates with *Candida* spp in the etiology and pathogenesis of denture stomatitis. The use of oral antimicrobial agents may provide a beneficial effect for denture stomatitis patients that are under antifungal therapy and that have poor oral hygiene and unfavorable salivary parameters.

Key words: Dental prosthesis; antifungal agents; *Candida*; *Streptococcus mutans*

Resumo

Objetivo: Verificar o número de *Streptococcus mutans* em saliva de pacientes com estomatite protética antes e após a terapia antifúngica.

Metodologia: Após exame clínico de 93 pacientes, 47 foram selecionados para exame micológico e desta amostra foram selecionados trinta pacientes: 15 com diagnóstico positivo e 15 com diagnóstico negativo de candidose foram avaliados para contagem de *S. mutans*, determinação de fluxo salivar e capacidade tampão. Higiene bucal e da prótese, tempo de confecção, tipo de lesão e dados salivares foram relacionados com características clínicas e laboratoriais de *Candida*.

Resultados: As lesões frequentes foram dos tipos I (43,5%) e II (53,5%). A quantidade de *S. mutans* foi seis vezes maior em pacientes com candidose e foi associada com baixo fluxo salivar e higiene oral deficiente. Após a terapia, a redução de *S. mutans* foi verificada particularmente em pacientes com fluxo salivar normal. Os valores variaram de 0,01 a 3,88 UFC/ml $\times 10^4$.

Conclusão: Os dados sugerem que os *Streptococcus* colaboram com *Candida* spp na etiopatogenia da estomatite protética. O uso de agentes antimicrobianos orais pode propiciar efeito benéfico para pacientes com estomatite protética submetidos à terapia antifúngica e que apresentam higiene oral deficiente e parâmetros salivares desfavoráveis.

Palavras-chave: Prótese dentária; agentes antifúngicos; *Candida*; *Streptococcus mutans*

Laurylene César de S. Vasconcelos^a
Fábio Correia Sampaio^a
Maria Carméli Correia Sampaio^b
Maria do Socorro Vieira Pereira^c
Maria Helena Pereira Peixoto^c

^a Department of Clinical and Social Dentistry, Federal University of Paraíba, João Pessoa, PB, Brazil

^b Department of Pharmaceutical Sciences, Federal University of Paraíba, João Pessoa, PB, Brazil

^c Department of Molecular Biology, Federal University of Paraíba, João Pessoa, PB, Brazil

Correspondence:

Laurylene César de Souza Vasconcelos
R. Maria Helena da Rocha, 113 - Aeroclub
João Pessoa, PB - Brasil
58036-823
E-mail: laurylene@uol.com.br

Received: May 20, 2009
Accepted: March 31, 2010

Conflict of Interest Statement: The authors state that there are no financial and personal conflicts of interest that could have inappropriately influenced their work.

Copyright: © 2010 Vasconcelos et al.; licensee EDIPUCRS. This is an Open Access article distributed under the terms of the Creative Commons Attribution-Noncommercial-No Derivative Works 3.0 Unported License.

Introduction

The surface of acrylic dentures usually host several microorganisms (fungi and bacteria) that are difficult to remove even by vigorous mechanical cleaning or chemical agents (1). This environment produced by the prosthesis can promote yeast proliferation with or without predisposing local factors (2).

Denture stomatitis is an oral pathology of multifactorial etiology that affects a large number of patients using complete or partial dentures. The main etiologic factors related to denture stomatitis are trauma, poor oral hygiene and infection with *Candida species*. In general, the patients with denture stomatitis complain of edema, hyperemia, pain in the affected areas and burning mouth when a low salivary flow is taking place (3,4).

The adhesion of *Candida* cells to oral surfaces is a key initial event in pathogenesis of oral candidiasis, and the complex structure of oral flora with mixed species communities can intermediate the predisposition for many oral conditions including candidiasis (3). There are evidences that denture stomatitis is not a result of *C. albicans* solely, but rather it is an outcome of multispecies biofilms that may include *Streptococcus mutans* and *Staphylococcus aureus* (5,6). In fact, it has been already observed that co-adhesion between *C. albicans* and many Streptococcal spp. promotes oral colonization by yeast cells (6). *Streptococcus mutans* is a frequent member of acrylic dentures surfaces and if incubated simultaneously with *Candida albicans* may compete for bidding sites but it can also promote yeast adhesion (5). This bacteria-yeast interaction has been a matter of investigation of many *in vitro* and *in vivo* studies (5,7-10). The analysis of the *in vitro* adherence of *S. mutans* and *C. albicans* can contribute to the understanding of the behavior of these organisms in the dental plaque. The interaction of these microorganisms in a combined culture can be understood as mutualistic, once both seem to be favored (11).

The adhesion is regarded as the first step for oral biofilm formation and distinct mechanisms of adherence can contribute for candidiasis resistance to antifungal therapeutic agents (8). In complex biofilms such as those found in the oral cavity, the ability of yeast to agglutinate with bacteria can be mediated by species interaction within the biofilm as well as by external factors like saliva, oral hygiene and exposure to antimicrobial agents (8,9). Despite the *in vitro* evidences of the *Candida*-bacteria interaction, few studies have investigated the *S. mutans* in patients with candidiasis or yeast related diseases (10,12). The aim of this study was to assess the number of *Streptococcus mutans* in saliva of patients with diagnosis of denture stomatitis before and after antifungal therapy.

Methods

The study protocol was approved by the Local Ethical Committee of the Federal University of Paraíba, João Pessoa, PB, Brazil. Only patients who were assisted at

the dental clinic and signed the consent were selected for the research. The criterion for participating in the research was dental prosthesis users. The exclusion criteria were: patients with systemic predisposing or confounding factor (diabetes, hyperthyroidism, immune diseases, radiotherapy and chemotherapy treatments), use of medicines that could affect any *Candida* infection such as antidepressive, immunosuppressive and anti-bacterial agents (2).

A total of 93 patients were examined and forty seven presented clinical signs of denture stomatitis. The clinical signs were classified in three stages described by Newton as pin-point hyperemia, diffuse hyperemia and nodular hyperemia of the entire denture area (13).

Swabs were used for collecting material from prosthesis (P1) and palatine lesions (P2) of patients with candidiasis. The material was subsequently placed on CHROMagar *Candida* for culture, isolation and identification of fungi species. CHROMagar is a selective and differential medium that allows selective isolation of yeasts and simultaneously identifies (by color reactions and colony morphology) colonies of *C. albicans*, *C. tropicalis* and *C. Krusei* with a high degree of accuracy (14).

Mycological exams as described previously were carried out in this sub-sample (4). The presence of *Candida* infection was confirmed in 32 patients, but only 15 presented more than 20 cfu (Colony Forming Units), a level which characterizes infection (1). Finally, the sample was composed of 30 patients with denture stomatitis: 15 with candidiasis and 15 without candidiasis.

Antifungal therapy was applied for the 15 patients who were positive for candidiasis. The patients received a tube of a commercial antifungal medicine (Daktarin® gel oral, Janssen Cilag Farmacêutica, 40 g) and instruction for using 3 times a day for two weeks. They were instructed to use a sufficient amount of gel that could cover the affected areas as recommended by the manufacturer. Denture were cleaned each night with neutral soap and washed with tap water. Patients were strongly requested to avoid using dentures at night during the test period.

Before starting the therapeutic period, oral hygiene parameters and salivary test were carried out before the therapeutic period of 2 weeks. Mycological and microbiological tests were carried out twice: before the patients have started using the medicine and two days after finishing the therapeutic period. Oral hygiene and prosthesis hygiene parameters were collected according to the guidelines of Sakki et al. (15). Saliva parameters (stimulated salivary flow rate and buffer capacity) were collected as described by Krasse et al. (16). The patients were classified in three levels of salivary flow rate: a) hiposalivation = 0.7mL/min, b) intermediate salivary flow = 0.71 up to 1.0 mL/min and c) normal salivary flow when above 1.1 mL/min. For buffer capacity, the reference levels were considered as normal for pH above 5 and low for pH below or equal to 4. The pH measurements were determined using indicate paper but intermediate values were checked in pH electrode for final classification (below of above 5.5).

The *S. mutans* counting was carried out for all 30 patients at baseline and for 15 patients after using the antifungal medicine. Non-stimulated saliva samples were placed in Brain Heart Infusion (BHI, Difco) and later on MSB medium: mitis salivarius agar (Difco) supplemented with 440 mM sucrose, 39 mM potassium tellurite and 0.2 units/mL of bacitracin, as recommended by Gold et al. (17). All procedures were in duplicate.

Statistical analysis was carried out using SPSS (Statistical Package for Social Sciences) v.10.0. The assumptions of equality of variances and normal distribution of errors were checked for all numerical variables. Since the distribution of the errors was homogeneous, data were tested by Student's T test with a significance level of 5%.

Results

Most patients were female (n=24) with an age range between 27 and 69 years-old. Within this range, the most frequent observations were between 49 up to 59-years-old. The lesions types observed were type I (43.5%) and type II (53.5%). Type III composed 3% of the lesions.

Candida albicans was the predominant yeast isolated (86.6%) followed by *C. tropicalis*, *C. parapsilosis*, *C. glabrata* e *C. krusei*.

Table 1 shows the mean cfu/mL of *S. mutans* and *Candida* spp of all patients. Ten subjects who showed high counts of *Candida* in the prosthesis were positive for candidiasis and were also with high number of *S. mutans* in saliva.

Table 1. Mean cfu/mL of *S. mutans* and *Candida* spp of 30 patients (15 patients positive and 15 negative for *Candida*).

N	N of colonies of <i>Candida</i> spp		Candidiasis	<i>S. mutans</i> Percentile (50%)	<i>S. mutans</i> (n x 10 ⁴ cfu/mL)
	Prosthesis	Palate			
1	0	0	-	-	1
2	0	0	-	-	3
3	0	0	-	-	11
4	0	0	-	-	12
5	0	0	-	-	13
6	> 200	17	+	-	14
7	0	0	-	-	16
8	0	0	-	-	16
9	103	> 200	+	-	17
10	>200	0	+	-	17
11	0	0	-	-	23
12	0	0	-	-	24
13	>200	122	+	-	26
14	0	0	-	-	29
15	0	0	-	-	29
16	> 200	113	+	-	29
17	0	0	-	+	53
18	0	0	-	+	55
19	0	0	-	+	58
20	0	0	-	+	85
21	> 200	20	+	+	92
22	> 200	126	+	+	154
23	> 200	0	+	+	174
24	116	19	+	+	175
25	> 200	149	+	+	206
26	> 200	0	+	+	254
27	> 200	93	+	+	322
28	183	0	+	+	382
29	170	0	+	+	446
30	> 200	39	+	+	495

Table 2 shows the reduction in salivary *S. mutans* (cfu/mL) after the antifungal therapy. In percentages, the differences range from 3.8 up to 82% with a mean reduction of 45%. Table 3 presents the mean (SD) values (cfu/mL) of *S. mutans* of patients before the antifungal therapy (baseline) according to other parameters. Table 4 presents the distribution of patients categorized in low and high *S. mutans* counting (cfu/mL) of patients who with and without candidiasis. The estimate amount *S. mutans* of in saliva among patients with deficient hygiene was ten fold higher compared to those with good hygiene. Among those patients with deficient hygiene (n=13), the mean (SD) values (cfu/mL) of *S. mutans* were 2.24(1.50) and 0.93(0.50) before and after the antifungal therapy, respectively. For those with good hygiene (n=2) there values were 0.21(0.10) and 0.12(0.01), respectively.

Table 2. Mean, standard deviation and confidence interval of *S. mutans* (cfu/mL) in saliva of patients (n=15) before and after the use of antifungal medicine.

N	Antifungal therapy		Difference (n x 10 ⁴ cfu/mL)
	Before (baseline) (n x 10 ⁴ cfu/mL)	After (n x 10 ⁴ cfu/mL)	
Mean	1.97	0.83	1.14
Standard deviation	1.56	0.57	1.27
Confidence interval	1.10-2.83	0.51-1.14	0.43-1.84

Table 3. Mean (SD) of fcu/mL of *S. mutans* (baseline), time of use of prosthesis, salivary flow rate, buffer capacity of patients who with and without candidiasis.

Candidiasis	N	<i>S. mutans</i> (n x 10 ⁴ cfu/mL)	Period of using prosthesis (months)	Salivary flow rate (mL/min)	Buffer capacity (pH)
Negative	15	0.3 (0.22) ^a	69.1 (49.3) ^a	1.2 (0.3) ^a	6.6 (0.7) ^a
Positive	15	1.97 (1.56) ^b	95.6 (83.5) ^b	0.7 (0.2) ^b	5.2 (1.2) ^b

Means in the same column followed by distinct superscripts indicate statistical significance (Student T test, P<0.05).

Table 4. Relationship between patients who showed low or high *S. mutans* in saliva and patients diagnosed as *Candida*-positive (presence of *C. albicans* on prosthesis or palate).*

<i>S. mutans</i> (n x 10 ⁴ fcu/mL)**	Diagnosis of candidiasis		Total
	Negative	Positive	
Low	11	5	16
High	4	10	14
Total	15	15	30

* Chi-square = 4.82, d.f.=1, p<0.05.

** cut-off point of 29 x 10⁴ cfu/mL of *S. mutans* (percentile 50%).

Discussion

C. albicans was the major isolated yeast in the selected subjects. This was also observed in other studies (2,9). The predominant site for colonies was the prosthesis indicating the importance of the mechanism of adherence of yeast cells on a non-shedding surface (6,15). Adhesion is the first step for yeast colonization on the oral epithelium, and dentures may become an important source for reinfection (18). For denture stomatitis patients the risk of candidiasis is high since porous acrylic surfaces can provide more bidding sites than natural teeth and shedding surfaces as oral mucosa (1).

Shinada et al. (8) observed that the presence of *Candida* was highest on the mucosal denture surfaces followed by clasp, tongue, and remaining teeth. Therefore, it is not a surprise that a significant number of patients could harbor *Candida albicans* in the prosthesis as shown in Table 1. From the clinical point of view an endogenously re-colonization of *Candida* in some patients cannot be disregarded. *Candida* is usually ubiquitous in the oral mouth and digestive tract. Thus a re-infection can be originated from external and internal sources, or both. Age of the prosthesis, oral hygiene and ability to penetrate the oral mucosa are certainly important variables to be considered for those less successful cases (1,2). How far oral hygiene and the counting of

S. mutans can influence re-infection, this is still a matter of debate (7).

Recently, many investigations have focus more on biofilm formation than in planktonic cells to explain the bacterial behavior in the *in vivo* oral environment (19). Regarding candidiasis, few studies have investigated the bacteria-yeast relationship using clinical parameters that can modulate the biofilm formation on different surfaces (5,10,12). It is important to consider bacterial-Candida interactions in the study of fungal virulence. Aspects of the host's microbial community might normally protect against fungal proliferation or might play an important role in the preparation of the fungus for its role in the infection. Also, mixed bacterial-fungal infections might have properties that are distinct from single-species infections (20). These co-species communities, probably existing as a mixed biofilm, are difficult to treat with antibiotics and antifungals (6).

Saliva and oral hygiene can be important mediators for oral biofilm development. Specific salivary components may act promoting yeast-bacteria co-aggregation and adherence to oral surfaces (9,4,17). Low salivary flow can also facilitate biofilm development and this is certainly a matter of concern when treating old subjects prone to dry mouth related problems (8,17).

In our study, *Candida* spp cfu values were obtained from biofilm material of dentures or palatal lesions, whereas *S. mutans* cfu levels were evaluated from saliva samples as an indication of *S. mutans* counting. Although *S. mutans* are major constituents of cariogenic oral biofilms, the saliva counting can be a good indicator of the major oral biofilms in the mouth in many oral conditions (21,22). In fact, a variation of *S. mutans* cells in different sites of the mouth is expected. Therefore, the option of collecting the *S. mutans* data from saliva was regarded as appropriate. In addition, all procedures were carried out following established guidelines of material collection (16). In any case, considering the cfu counting of *S. mutans* in the evaluated subjects, one must estimate that in denture oral biofilms there are at least 5-fold more bacteria than those found in saliva. Girard Junior et al. (2) observed that the dental biofilm formed in the presence of sucrose would be an indirect contributing factor for yeast adherence due to an increase in *Streptococcus* spp. As shown in Tables 1 and 4, most of the subjects with high counts of *Candida* also had high counts of *S. mutans*. Moreover, subjects with candidiasis and high amounts of *S. mutans* in

the oral mouth were the virtually the same with unfavorable salivary parameters and poor oral hygiene. The question at this point is not if a potentialization of the adherence capacity of *Streptococcus mutans* and *Candida albicans* takes place in a biofilm development, but rather if this synergic effect can be estimated or predicted clinically. The clinical implications of these results are that candidiasis treatment can be more effective when including frequent oral hygiene measures to reduce biofilm formation and *S. mutans* in the oral mouth (23). On the other hand, a risk for dental caries due to *Candida* is likely as reported by Moalic et al. (24) and Nikawa et al. (25). A low biofilm pH may favor *S. mutans* but *Candida* can also take advantage of adherence mechanism in such environment.

The observation of lesions types I and II in these subjects indicates that candidiasis was not an acute event in these patients. Poor oral hygiene and unfavorable salivary parameters can probably aggravate this condition by many ways including the *Candida-Streptococcus* co-aggregation. From the clinical point of view, compliance for oral health instructions can be problematic. Therefore, in addition to oral hygiene instructions, the use of antimicrobials might be an interesting choice to patients with a high *S. mutans* counting and resistant candidiasis.

Conclusions

It can be concluded that:

1. *S. mutans* counting was higher in patients with candidiasis and it was associated with low salivary flow and poor oral hygiene;
2. After therapy, it was verified a reduction of *S. mutans* particular in patients with normal salivary parameters; and
3. *Streptococcus* spp collaborates with *Candida* spp in the etiology and pathogenesis of denture stomatitis. The use of oral antimicrobial agents may provide a beneficial effect for denture stomatitis patients that are under antifungal therapy and that have poor oral hygiene and unfavorable salivary parameters.

Acknowledgments

This study is part of a doctoral thesis and it was supported by Lauro Wanderley University Hospital for its realization.

References

1. Budtz-Jørgensen E. Etiology, pathogenesis, therapy and prophylaxis of oral yeast infections. *Acta Odontol Scand* 1990;48:61-9
2. Girard Junior B, Landry RG, Giasson L. La stomatite prothétique: étiologie et considérations cliniques. *J Can Dent Assoc* 1996;62:808-12.
3. Könsberg R, Axéll T. Treatment of Candida - infected denture stomatitis with a miconazole lacquer. *Oral Surg Oral Med Oral Pathol* 1994;78:306-11.
4. Vasconcelos LC, Sampaio MC, Sampaio FC, Higino JS. Use of Punica granatum as an antifungal agent against candidosis associated with denture stomatitis. *Mycoses* 2003;46:192-6.
5. Linossier A, Vargas A, Villegas R, Chimenos E. Quantitative relationship between salivary level of *Streptococcus mutans* and *Candida albicans* in children with Down's syndrome. *Med Oral* 2002;7:284-92.

6. Baena-Monroy T, Moreno-Maldonado V, Franco-Martinez F, Aldape-Barrios B, Quindos G, Sanchez-Vargas LO. *Candida albicans*, *Staphylococcus aureus* and *Streptococcus mutans* colonization in patients wearing dental prosthesis. *Med Oral Patol Oral Cir Bucal* 2005;11:E27-39.
7. Holmes AR, Gopal PK, Jenkinson HF. Adherence of *Candida albicans* to a cell surface polysaccharide receptor on *Streptococcus gordonii*. *Infect Immun* 1995;63:1827-34.
8. Shinada K, Ozaki F, Cordero JG, Okada S, Shimoyama K, Nagao M, Ichinose S, Yamashita Y. A morphological study of interactions of *Candida albicans* and *Streptococcus mutans*. *Kokubyo Gakkai Zasshi* 1995;62:281-6.
9. Jorge AOC, Koga-Ito CY, Gonçalves CR, Fantinato V, Unterkircher CS. Presença de leveduras do gênero *Candida* na saliva de pacientes com diferentes fatores predisponentes e de indivíduos controle. *Rev Fac Odontol Univ São Paulo* 1997;11:279-85.
10. El-Azizi MA, Starks SE, Khardori N. Interactions of *Candida albicans* with other *Candida* spp. and bacteria in the biofilms. *J Appl Microbiol* 2004;96:1067-73.
11. Barbieri DSV, Vicente VA, Fraiz FC, Lavoranti OJ, Svidzinski TIE, Pinheiro R L. Analysis of the *in vitro* adherence of *Streptococcus mutans* and *Candida albicans*. *Braz j Microbiol* 2007;38:624-31.
12. Shinada K, Teraoka K, Asaka T, Cordeiro JG, Ozaki F, Shimoyama K et al. Distribution of *Candida* species and *mutans streptococci* related to oral conditions in elderly persons. *Kokubyo Gakkai Zasshi* 1997;64:512-17.
13. Newton AV. Denture Sore Mouth: a possible Aetiology. *Br Dent J* 1962;357-60.
14. Pfaller MA, Houston A, Coffmann S. Application of CHROMagar *Candida* for rapid screening of clinical specimens for *Candida albicans*, *Candida tropicalis*, *Candida krusei* and *Candida (Torulopsis) glabrata*. *J Clin Microbiol* 1996;34:58-61.
15. Sakki TK, Knuutila MLE, Läärä EL, Anttila SS. The association of yeasts and denture stomatitis with behavioral and biologic factors. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 1997;84:624-9.
16. Krasse B. Risco de cárie: um guia prático para avaliação e controle. 2ª ed. São Paulo: Quintessence Books; 1988.
17. Gold OG, Jordan HV, Van Houte J. A selective medium for *Streptococcus mutans*. *Arch Oral Biol* 1973;18:1357-64.
18. Noborikawa E, Silveira FRX, Witzel, AL, Lotufo, MA. Biotypes of *Candida albicans* isolates from the oral mucosa of HIV seropositive and control subjects. *Rev Odonto Ciênc* 2009;24:258-63.
19. Marsh PD. Role of the oral microflora in health. *Microb ecol health dis* 2000;12:130-7.
20. Wargo MJ, Hogan, DA. Fungal-bacterial interactions: a mixed bag of mingling microbes. *Curr Opin Microbiol* 2006;9:359-64.
21. Sullivan A, Borgström MK, Granath L, Nilsson G. Number of *mutans streptococci* or *lactobacilli* in a total dental plaque sample does not explain the variation in caries better than the numbers in stimulated whole saliva. *Community Dent Oral Epidemiol* 1996;24:159-63.
22. Hintao J, Teanpaisan R, Chongsuvivatwong V, Ratarasan C, Dahlen G. The microbiological profiles of saliva, supragingival and subgingival plaque and dental caries in adults with and without type 2 diabetes mellitus. *Oral Microbiol Immunol* 2007;22:175-81.
23. Vasconcelos LCS, Sampaio FC, Sampaio MCC, Pereira MSV, Higino JS, Peixoto MHP. Minimum inhibitory concentration of adherence of *Punica granatum Linn* (pomegranate) gel against *S. mutans*, *S. mitis* and *C. albicans*. *Braz Dent J* 2006;17:223-7.
24. Moalic E, Gestalin A, Quinio D, Gest PE, Zerilli A, Le Flohic AM. The extent of oral fungal flora in 353 students and possible relationships with dental caries. *Caries Res* 2001;35:149-55.
25. Nikawa H, Egusa H, Makihira S, Nishimura M, Ishida K, Furukawa M, Hamada T. A novel technique to evaluate the adhesion of *Candida* species to gingival epithelial cells. *Mycoses* 2003;46:384-9.