Natural killer activity of the spleen cells of Ehrlich tumor-bearing mice treated with Copaifera multijuga extract


ABSTRACT

Aims: Copaifera multijuga Hayne oleoresin is commonly used in traditional medicine owing to its anti-inflammatory, antiseptic, antitumor, and antibacterial properties. However, little is known about the effect of the compounds from the bark of this plant. In this study, the immunomodulatory effect of the ethanolic extract of C. multijuga bark via natural killer activity of non-adherent spleen cells of Ehrlich tumor-bearing mice was evaluated.

Methods: Male Swiss mice were inoculated subcutaneously with 1×10^6 Ehrlich tumor cells (Ehrlich and Ehrlich/C. multijuga group) or phosphate buffered saline solution (control group and C. multijuga group) and treated orally daily with C. multijuga extract (200 mg kg^{-1}, 0.1 mL per mouse, for the Ehrlich/C. multijuga and C. multijuga groups) or phosphate buffered saline solution (control group and Ehrlich group). The four experimental groups consisted in eight mice each and were organized in two sets, one treated for seven days and another treated for 14 days, totaling 64 mice throughout the experiment. Twenty-four hours after the last oral administration, the mice were euthanized and the spleen tissue was isolated to prepare a non-adherent spleen cell suspension and to evaluate natural killer activity. Data are presented as the cell lysis percentage of Yac.1 target cells by non-adherent spleen cells.

Results: Treatment for seven days increased natural killer activity in the Ehrlich/C. multijuga group (21.20±8.89, p<0.05) compared to the control group (3.14±2.71, p>0.05); however, this effect was not maintained in the groups treated for 14 days (Control: 6.02±6.98, Ehrlich: 4.82±7.7, C. multijuga: 2.07±2.10, Ehrlich/C. multijuga: 2.01±1.63, p<0.05).

Conclusions: Treatment for seven days with an ethanolic extract of C. multijuga bark enhanced the natural killer activity of non-adherent spleen cells from Ehrlich tumor-bearing mice.

Keywords: Copaifera multijuga; fabaceae; copaiba; natural killer cells; carcinoma, Ehrlich tumor; immunomodulation.
INTRODUCTION

Phytotherapeutic compounds have been widely used due to their efficiency, low toxicity, biocompatibility and low cost [1]. Since 2002, the World Health Organization has recognized the importance of phytotherapeutic compounds used in traditional medicine as part of conventional healthcare strategies [2]. In Brazil, the National Policy of Medicinal Plants and Herbal Medicines encourages the development of plant-based compounds for use in public health programs [2].

In this context, copaiba has been one of the most important Brazilian medicinal plants [1-6]. As early as 1587, the Portuguese explorer and naturalist Gabriel Soares de Sousa described the use of copaiba oleoresin for healing arrow wounds [2]. Nowadays, copaiba is used in medicine owing to its anti-inflammatory, antibacterial, antifungal, leishmanicidal, antitumor, and analgesic properties [1-5, 7-9].

Copaiba trees belong to the genus Copaifera, family Fabaceae, and subfamily Caesalpinioideae [6]. There are more than 70 Copaifera species distributed worldwide, with widespread occurrence in Central and South America, but also in West Africa and Asia [6]. Brazil has the greatest biodiversity of Copaifera with 26 species and 8 varieties [6]. The most abundant species are Copaifera multijuga Hayne, Copaifera reticulata, and Copaifera langsdorffii Desf. [5-7].

The oleoresin is the most important economic and medicinal product of copaiba trees [6]. It is a transparent and colored exudate obtained from the trunks of the trees, with variable viscosity, and consisting of a nonvolatile fraction of diterpenes (20%) and a volatile fraction of sesquiterpenes (80%) [2, 5-7]. Copaiba oleoresin chemical profile may vary according to species, seasonal and climatic characteristics, soil type and composition, and biotic pressures, such as insect predation and pathogen infection [2, 5-7]. The production of oleoresin per tree ranges from 100 mL to 60 L annually; however, not all trees produce oil [6]. Therefore, several parts and preparations of the plant, such as stem bark crude formulation, are used in folk medicine [6].

In previous studies by our group, it was observed that ethanolic extract of C. multijuga bark can affect Ehrlich tumor cells, reducing their viability in vitro as well as their development in vivo [9]. Ehrlich tumor is a spontaneous murine mammary adenocarcinoma adapted to the ascitic form and inoculated in mice by serial intraperitoneal passages [9, 10]. This tumor model is widely used in experimental cancer studies due to its versatility; it is able to adapt to an ascitic form when inoculated by the intraperitoneal route or a solid form when inoculated by the subcutaneous route [9, 11, 12].

The development of Ehrlich ascites tumor is commonly accompanied by intense alterations of the immune response, leading to decreased immunocompetence with down-regulation of cytotoxic cells such as natural killer (NK) cells [13]. NK cells are important effector cells of the innate immune system that play an active role in the elimination of cancerous and virus-infected cells [14]. Activation of NK cells results in their lytic granule exocytosis, with the release of perforin and granzymes against locally attached target cells; this leads to the secretion of cytokines [13, 14]. An important feature of the NK immune response is that these cells do not require prior sensitization to exert their effector function [14].

Thus, considering the antitumor effect of C. multijuga bark extract on Ehrlich tumor [9] and the importance of NK cells as a strategy in cancer immunotherapy [15], the present study aimed to evaluate the immunomodulatory effect of ethanolic extracts obtained of C. multijuga bark on the NK activity of the non-adherent spleen cell culture of Ehrlich tumor-bearing mice.

METHODS

Plant material

The plant material was collected at a particular property in Guarantã do Norte, MT, Brazil (S 9°48’ 31.0” W 54°53’ 18.0”), according to Albiero et al. [9]. The specimens were identified by Professor Ivani Kuntz Gonçalves, and samples are deposited at Herbarium of Federal University of Mato Grosso (Universidade Federal de Mato Grosso, UFMT), Biological Collection of Southern Amazonia (Acervo Biológico da Amazônia Meridional, ABAM), Sinop, MT, Brazil, N.4801.

Preparation of C. multijuga extracts

C. multijuga extracts were prepared according to Albiero et al. [9]. C. multijuga bark was obtained at a
stem depth of 5 cm, cleaned, cut in small pieces, and air-dried at 40 °C for seven days. The dried material was processed in a crusher (1.0 mm) and macerated with a different solvent for each extraction phase (2 L per 1,257 g sample). First, hexane was added and the extract was stored for seven days. Ethyl acetate was added, and it was stored for an additional seven days. Ethanol was added, and the extract was stored for seven days to complete the maceration. At each phase, the solvent was replaced, and the extract was protected from light exposure. The resulting hexane, ethyl acetate, and ethanolic extracts were filtered through a filter paper under negative pressure. A rotary vacuum evaporator (IKA® RV 05 basic, Staufen, Germany) at 40 °C and vacuum desiccator were used to concentrate and remove the solvents. The yields obtained were 1.652 g for the hexane extract, 5.152 g for the ethyl acetate extract, and 220.463 g for the ethanol extract. The ethanolic extract was used for the evaluation of NK cell activity in vitro, since it obtained the highest yield and the best result with in the in vitro cytotoxicity analysis demonstrated in previous study [9].

**Animals**

Male Swiss mice, aged 40 to 50 days, were obtained from the Central Animal Facility of UFMT in Cuiabá, MT, Brazil. The mice were housed in polypropylene boxes at 22 °C, exposed to 12/12-h light/dark cycles, and administered filtered water and pelleted feed (Purina, St. Louis, Missouri, USA) ad libitum. All procedures were approved by the Ethics Committee on Animal Use of UFMT (Protocol no. 23108.700603/14-3).

The mice were inoculated subcutaneously with 1×10^6 tumor cells (Ehrlich or Ehrlich/C. multijuga groups) or 100 μL of phosphate buffered saline (PBS) (Control or C. multijuga groups). Twenty-four hours later, the mice were treated daily by gavage with the ethanolic extract (200 mg kg⁻¹; 100 µL per animal; for the Ehrlich/C. multijuga and C. multijuga groups) or vehicle (PBS; 100 µL per animal; control and Ehrlich groups). The four experimental groups consisted in eight mice each and were organized in two sets, one treated for seven days and another treated for 14 days, totaling 64 mice throughout the experiment. After these periods, the mice were euthanized for the in vitro evaluation of the NK activity from the spleen cell culture.

**Spleen cell suspensions**

Spleen cell suspensions were obtained by teasing the spleens on a sterile fine nylon screen in Roswell Park Memorial Institute (RPMI) 1640 medium (Cultilab, Campinas, SP, Brazil). The cell suspensions were centrifuged at 1,500 rpm for 10 min and suspended in 1 mL of complete medium (RPMI 1640 supplemented with 20% of heat-inactivated fetal bovine serum (FBS) (Cultilab, Campinas, SP, Brazil).

**Ehrlich tumor cell suspension**

Ehrlich tumor was provided by Rondon Tosta Ramalho, Ph.D., from the Federal University of Mato Grosso do Sul, Campo Grande, Brazil and was maintained through intraperitoneal inoculation (ascitic form) in Swiss mice, every seven days. Tumor cell suspensions were prepared in sterile PBS, to final concentration of 1×10^7 viable cells mL⁻¹. The mice were inoculated subcutaneously in the right flank region (0.1 mL per animal). Viability, assessed by Trypan Blue dye exclusion method, was at least 70%.

**Yac.1 Target Cell Suspension**

Yac.1 mouse lymphoma cell line, an NK-sensitive tumor cell line, was provided by Rio de Janeiro Cell Bank. Aliquots of Yac.1 cells were cultured in complete medium at 37 °C and 5% CO₂ for seven days. The cultured suspended cells were centrifuged, resuspended in RPMI 1% FBS, and its concentration was adjusted to 1×10⁶ cells mL⁻¹.

**Colorimetric assay for cytotoxic activity analysis**

A non-radioactive colorimetric method based on lactate dehydrogenase (LDH) activity measurements was used to evaluate cytotoxic activity (Cytotoxicity Detection Kit, Roche Diagnostics, Mannheim, Germany). Mononuclear cells were obtained by centrifuging the spleen cell suspension on a Ficoll-Hypaque gradient (Sigma-Aldrich, St. Louis, United States of America), followed by incubation on glass Petri dishes for 90 min at 37 °C to remove the adherent cells. The non-adherent cells were gently recovered from the Petri dishes and resuspended in RPMI 1% FBS adjusted to a cell density of 1×10⁷ cell mL⁻¹. Next, 100 µL of the non-adherent cell suspension (effectors) was dispensed into a “U”-bottomed 96-well microtiter plate with 100 µL of the target cell suspension (Yac.1) at a concentration of 1×10⁶ cell mL⁻¹ (effector to target ratio 50:1). Maximal lysis of target cells was determined by adding 100 mL of Triton X solution (Sigma-Aldrich, St. Louis, United...
States of America). Spontaneous lysis of Yac.1 cells was determined by incubation with RPMI 1% FBS. Background control was also maintained with RPMI 1% FBS, without cells. After 4 h of incubation at 37 °C and 5% CO2, the plate was centrifuged for 10 min at 1500 rpm and 50 μL of the supernatant was carefully removed from each well and transferred into a 96-well flat-bottomed microtiter plate (Nunc A/S, Roskilde, Denmark). LDH activity was quantified using 50 μL of a Diaphorase/NAD+ mixture and a dye solution containing iodo tetrazolium chloride. Sodium lactate was added to each well and incubated for 30 min at room temperature away from light. The absorbance was read at 492 nm (Thermo plate TP-reader). The percentage of specific lysis was calculated according to the following formula: cytotoxicity (%) = \[\frac{\text{absorbance of the mixture of target and effector cells} - \text{absorbance of control effector cells}}{\text{absorbance of maximal lysis control} - \text{absorbance of spontaneous lysis control}}\] × 100. Data are presented as means ± standard deviation. Before cytotoxicity calculation, all absorbance values were subtracted from the mean value of the background control.

The cytotoxic assay of NK activity measures the ability of non-adherent spleen cells to kill Yac.1 target cells. This effect is attributed to NK cells because these cells are able to kill the Yac.1 target cells without prior contact [13, 14].

**Statistical Analysis**

Statistical analysis was performed utilizing the GraphPad Instat software (San Diego, California, USA). One-way analysis of variance (ANOVA) and Tukey-Kramer tests were employed. Differences were considered significant when the probability of error was lower than 5% (p≤0.05).

**RESULTS**

**Figure 1** presents the cytotoxicity of non-adherent spleen cell suspension from experimental groups of mice treated with the ethanolic extract for seven or 14 days. Treatment with the ethanolic extract of *C. multijuga* bark for seven days (Figure 1A) increased the NK activity in Ehrlich tumor-bearing mice (21.20±8.89) compared to the control group (3.14±2.71; p<0.05) and *C. multijuga* group (5.04±6.07; p<0.05). However, this effect was not maintained in the groups treated for 14 days (Figure 1B) (Control: 6.02±6.98, Ehrlich: 4.82±7.72, *C. multijuga*: 2.07±2.10, Ehrlich/*C. multijuga*: 2.01±1.63; p>0.05).

**DISCUSSION**

NK cells are important innate immune effector cells that provide rapid response to tumor and virus-infected cells [16]. Their importance is underscored by the fact that patients with NK cell deficiency suffer from severe recurring systemic and life-threatening infections [16]. Strikingly, the high cytotoxic activity of peripheral blood NK cells is associated with 10% lower incidence of tumors in men and 4% in women, and their infiltration of certain tumor tissues is an indicator of better disease prognosis [16]. In addition, NK cells that infiltrate tumors may prevent tumor metastasis and lymphatic invasion [13].
It is well known that the current treatments for cancer, such as chemotherapy and radiotherapy, induce immunosuppression in patients, decreasing immune function or causing dysfunction in immune cells such as cytotoxic T-lymphocytes and NK cells [14, 17]. NK cells or their precursors are highly sensitive to in vivo treatment with cyclophosphamide, a standard chemotherapeutic agent, with reduction of NK cell activity in mice [13]. In this way, promotion of NK cell activation and proliferation is an emerging strategy in cancer immunotherapy, mainly because NK cells can lyse tumor cells without prior activation [15]. Furthermore, NK cells produce and secrete potent immunoregulatory cytokines, particularly IFN-γ, which increases cell reactivity and activates macrophages against the tumor cells [18].

In the present study, the ethanolic extract of C. multijuga bark was used to stimulate the immune response of mice, enhancing or potentiating the host’s defense mechanism to inhibit tumor growth. Immunomodulation through natural substances may be considered as an alternative for prevention and cure of neoplastic diseases [17]. Immunomodulators may activate cytotoxic effector cells, such as cytotoxic T-lymphocytes, NK cells, and macrophages to kill the tumor cells without harming the normal host cells [14, 17].

Previous study by our group demonstrated that the ethanolic extract of C. multijuga bark can affect Ehrlich tumor cells, reducing their viability in vitro as well as their development in vivo, reducing tumor mass to 45%, and increasing the production of IL-12p70, TNF-α, and IFN-γ in Con A or SAC-stimulated spleen cell culture supernatants [9]. C. multijuga bark ethanolic extract also increased the antioxidant capacity of the liver, increased catalase, reduced glutathione and glutathione-S-transferase activities, and decreased the levels of lipid peroxidation [19]. Results of the present study corroborate previous data [9, 19], showing that the immunomodulatory effect of C. multijuga on Ehrlich tumor-bearing mice is also a consequence of NK cell activity stimulation.

The use of medicinal plants for therapeutic purposes is now spread worldwide [20]. Several herbal preparations used in the indigenous system of medicine are known to boost the immune system [14]. Within this context, C. multijuga is an interesting option because of its documented medicinal effects such antimicrobial, antinociceptive, anti-inflammatory, wound-healing, antifungal, antiparasitic, and antitumor properties [1, 5, 7-9, 20]. These activities are attributed to the presence of sesquiterpene and diterpene metabolites [20, 21].

Although studies associating the biological response of C. multijuga sesquiterpene and diterpene metabolites with the activity of NK cells are lacking, some authors described that the activity of NK cells can be improved by sesquiterpenes from Artemisia annua L. [15], Vernonia cinerea L. [14, 17], and Zanthoxylum rhoifolium L. [2].

Artemisinin is a sesquiterpene lactone extracted from the plant of sweet wormwood (A. annua L) and is a Chinese traditional medicine that has been used in the treatment of malaria [15]. Artemisinin also causes the apoptosis of various cancer cells such as those of the colon, breast, lung, and pancreas, because it reacts with heme or free iron, generating cytotoxic radicals that induce oxidative damage in these cells [15]. Houth et al. [15] demonstrated that artemisinin significantly enhances NK cell activity through granule exocytosis via the stimulation of signaling molecules of NK cell activating receptor.

Silva et al. [18] investigated the antitumor properties of the volatile oil from Z. rhoifolium leaves and some terpenes (α-humulene, β-caryophyllene, α-pyrene, and β-pyrene) in vitro and in vivo using the Ehrlich ascites tumor model. The volatile oil had significant activity against the Ehrlich ascites tumor at a dose of 20 mg kg⁻¹, increasing survival of tumor-bearing mice to 80%; whereas survival of mice treated with 20 mg kg⁻¹ of β-caryophyllene presented an increase of 31% [18]. The same response profile was observed, with half maximal inhibitory concentration (IC₅₀) values of 37 μg mL⁻¹ for volatile oil and 102 μg mL⁻¹ for β-caryophyllene [18]. The authors also demonstrated that treatment of tumor-bearing mice with 20 mg kg⁻¹ of volatile oil or β-caryophyllene induced higher levels of splenic NK cell activity [18].

Vernolide-A is a sesquiterpene lactone present in the plant V. cinerea, which has many therapeutic uses in traditional medicine worldwide [14, 17]. Its antioxidant, immunomodulatory, chemoprotective, and radioprotective effects in mice have been reported [14, 17]. Administration of vernolide-A enhanced NK cell activity, as well as augmented antibody-dependent cellular cytotoxicity and antibody-dependent complement-mediated cytotoxicity in Ehrlich ascites tumor-bearing mice [17] and in B16F-10 metastatic melanoma-bearing mice [14]. In both situations, the increase in NK cell activity was associated with the up-regulation of IL-2 and IFN-γ cytokines [14, 17].

The studies of Silva et al. [18], Pratheeshkumar and Kuttan [14, 17], and Houth et al. [15] support the
enhancement in the NK activity from the culture of non-adherent spleen cells of Ehrlich tumor-bearing mice after treatment with the ethanolic extract of *C. multijuga* bark observed in this study; increased pro-inflammatory cytokines, such as IFN-γ, was shown to be the mechanism in a previous study [9]. IFN-γ is produced by T lymphocytes, NK cells, macrophages, and neutrophils and has receptors on virtually all cell types in the body [17]. Increased production of IFN-γ is typically associated with an effective host defense against intracellular pathogens and cancer [17].

Immunomodulatory effects are also attributed to phenolic compounds and flavonoids [22-24]. Pereira et al. [25] evaluated the antioxidant effects of *C. multijuga* bark ethanolic extract and observed a higher concentration of phenolic compounds, such as epicatechin and epiafzelechin-condensed tannins. Increased NK activity observed in this study may be a consequence of the condensed tannins present in *C. multijuga* extract since condensed tannins upregulate NK cell response [22, 23].

Although the improvement in NK activity seems to be opposite to the anti-inflammatory effect of copaiba, it is important to note that the biological effect of the plant extract reflects the complexity of the compounds and their different mechanisms of action [9, 26]. Thus, reduction of Ehrlich tumor growth may be a result of the association of the anti-inflammatory and immunostimulatory effects.

In fact, during the Ehrlich tumor growth, the progressive reduction of NK cell activity was related to the presence of prostaglandin E2 [13]. In addition, copaiba extracts appear to be similar to non-steroidal anti-inflammatory drugs, involving inhibition of cyclooxygenase and lipoxygenase pathways [9, 27]. Thus, the extract inhibits the influx of neutrophils and macrophages into the tumor microenvironment, reducing the angiogenic factors important for tumor growth, as well as increasing the antitumor immune response, improving the NK cells activity and reducing the immunosuppressive factors such as prostaglandins [9, 13, 27].

An ideal anti-cancer drug should show killing activity only against cancer cells with no toxic effects on normal and immune system cells [15].

In this respect, compounds from *C. multijuga* may be ideal, owing to their high effectiveness and low toxicity. Albiero et al. [9] showed that ethanolic and ethyl acetate extracts from *C. multijuga* bark are not toxic to normal spleen cells *in vitro* at 0.5 mg mL⁻¹ and 0.25 mg mL⁻¹ concentrations. Furtado et al. [28] showed no genotoxic effects in both *in vitro* and *in vivo* micronucleus assays using different concentrations of *C. multijuga* leaf extracts and oleoresin. No genotoxic effects were also observed by Alves et al. [29] in their micronucleus tests with *C. multijuga* oleoresin.

Thus, the results of this study corroborate with previous reports regarding the effects of copaiba, showing that the compounds may be potent anticancer drugs that act not only to inhibit cancer development but also to activate antitumor immune response. Although oleoresin is the most used form of copaiba, it is important to consider that not all trees produce the oleoresin [6], and other forms, such extracts from the bark, may also be used since they also have bioactive compounds.

It can be concluded that the NK activity from the culture of non-adherent spleen cells of Ehrlich tumor-bearing mice was enhanced by treatment with the ethanolic extract of *C. multijuga* bark for seven days. Additional studies are necessary to identify which compounds may have produced this effect and which cellular mechanisms are involved.

NOTES

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**Conflicts of interest disclosure**

The authors declare no competing interests relevant to the content of this study.

**Authors’ contributions**

All the authors declare to have made substantial contributions to the conception, or design, or acquisition, or analysis, or interpretation of data; and drafting the work or revising it critically for important intellectual content; and to approve the version to be published.

**Availability of data and responsibility for the results**

All the authors declare to have had full access to the available data and they assume full responsibility for the integrity of these results.

REFERENCES


