

# EVALUATION OF DIMENSIONAL STABILITY OF IMPRESSION MATERIALS IMMERSSED IN DISINFECTANT SOLUTIONS USING A METAL TRAY

## AVALIAÇÃO DA ALTERAÇÃO DIMENSIONAL DE MATERIAIS DE MOLDAGEM IMERSOS EM SOLUÇÕES DESINFETANTES USANDO UMA MATRIZ METÁLICA

Panza, Leonardo Henrique Vadenal\*  
 Porto, Vinicius Carvalho\*\*  
 Salvador, Milton Carlos Gonçalves\*\*\*  
 Silva Rosa, Odila Pereira da\*\*\*\*

### ABSTRACT

**Introduction:** When impressions are taken, saliva and blood are removed from the material, and washing them does not always guarantee that the material is completely clean. Therefore, methods for disinfecting impressions (immersing in solutions) are a necessity, but they can affect the accuracy of dental impressions. **Objective:** This study aimed to evaluate the dimensional stability of dental impression materials immersed in disinfectant solutions. **Material and methods:** This study used 45 impressions of each of the following materials Impregun F® (polyether) and Hydrogun® (irreversible hydrocolloid). Sodium hypochlorite and glutaraldehyde were selected as disinfectants and the immersion times were 10 and 15 minutes. Impressions of each material were immersed in both solutions: 2% glutaraldehyde solution (Milton 1%) and sodium hypochlorite solution (Milton 1%), for 10 and 15 minutes. Impressions for 15 min. The control group without immersion in disinfectant solutions. **Results:** Neither group showed any statistically significant difference (ANOVA) in the measurements after being soaked in the two disinfectant solutions. However, impressions were disinfected by sodium hypochlorite for 15 minutes showed a dimensional change (~0,122 mm) compared with control group. **Conclusion:** This study it can be concluded that the immersion practices for disinfecting impressions affect the quality of impressions obtained, except when sodium hypochlorite was used and immersion time was 15 minutes.

**UNITERMS:** dimensional stability, impression materials.

### SUMO

**Introdução:** Quando se tomam impressões de modelos para confecção de próteses, principalmente que são tomadas em áreas retentivas da boca, resíduos de saliva e sangue ficam retidos no material. Portanto, métodos de desinfecção das impressões durante que todo sangue e microorganismos aderidos à superfície do molde são necessários. Assim torna-se necessária a desinfecção dos moldes para a qual os métodos de desinfecção spray e imersão têm sido testados com várias soluções desinfetantes. **Objetivo:** Este estudo objetivou avaliar a estabilidade dimensional dos materiais de moldagem após a

\* Graduated student, UNICAMP, Piracicaba Dental School.

\*\* Assistance Professor of Prosthodontics Department, University of São Paulo, Bauru Dental School.

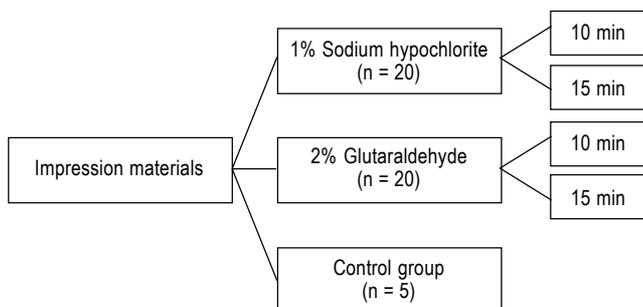
\*\*\* Associated Professor of Prosthodontics Department, University of São Paulo, Bauru Dental School.

\*\*\*\* Associated Professor of Microbiology Department, University of São Paulo, Bauru Dental School.



Forty-five impressions were made with each of the following materials: Impregun F® (polyether), Permlastic® (polysulfide) and Hydrogun® (irreversible hydrocolloid). The selected disinfectants were: 1% sodium hypochlorite (Milton) and 2% glutaraldehyde (Glutaacid®). They were freshly prepared for each experiment.

The experimental groups were divided according to the diagram:



Ten impressions from each group of materials were immersed in glutaraldehyde solution for 10 min, and 10 impressions of the same materials were immersed for 15 min. The same procedure was carried out with the sodium hypochlorite solution. Five (5) impressions of each material were used as control group, without being immersed in any disinfectant solution.

The impression materials were dispersed and mixed according to the manufacturer's recommendations at room temperature (23°C). A gradual, constantly increasing pressure was applied to a perforated metal cast in order to avoid excess material. Afterwards, a 1 mm thick layer was placed over the cast and the impression was

separated from the tray after 6 minutes. They were measured immediately after the impression procedure, to prevent any risk of distortion. Next, every impression was immersed in a disinfectant solution for 10 or 15 minutes, removed and rinsed under cold running water for 30 seconds and dried with compressed air. Immediately after drying, the original impressions were magnified × 30 and the distances between the lines AB, CD, AC and BD as shown in Figure 1 were measured three times by two examiners, for each elastomeric material. Irreversible hydrocolloid impressions were measured twice, because this material is more susceptible to syneresis and drench (Andir et al.<sup>2</sup>, 2002). Mitutoyo digital measuring microscope (TM500) with a sensitivity of 1.0 μm was used.

The analysis of variance (ANOVA) model was used to test if there was no difference between the disinfectants and the control, and if there was no difference between the impression material after 10 and 15 minutes. The hypotheses tested were: H0: no difference between the groups. H1: difference of confidence.

Forty-five impressions were made and divided into three groups: control and experimental groups. The graphic presentation of the data in Table 1 shows the means and standard deviations. The standard deviations ranged from 0.009 to 0.140 for alginate impressions, 0.004 to 0.092 for polyether and 0.016 to 0.149 for polysulfide. The results of the measurements obtained are expressed in millimeters.

TABLE 1 – means and standard deviations.

Material/Control	Means/AB	Means/CD	Means/AC	Means/BD
Alginate/control	24,062 ± 0.035	24,108 ± 0.053	4,197 ± 0.009	4,094 ± 0.016
Alginate/Hypochlorite 10 min	24,008 ± 0.046	24,077 ± 0.140	4,208 ± 0.029	4,108 ± 0.044
Alginate/Hypochlorite 15 min	23,871 ± 0.123	23,804 ± 0.122	4,191 ± 0.035	4,182 ± 0.095
Alginate/Glutaraldehyde 10 min	23,945 ± 0.126	23,935 ± 0.094	4,165 ± 0.047	4,079 ± 0.049
Alginate/Glutaraldehyde 15 min	23,983 ± 0.090	23,982 ± 0.098	4,166 ± 0.040	4,071 ± 0.024
Polyether/Control	23,997 ± 0.092	24,049 ± 0.056	4,182 ± 0.052	4,125 ± 0.045
Polyether/Hypochlorite 10 min	24,061 ± 0.021	24,079 ± 0.029	4,204 ± 0.007	4,139 ± 0.004
Polyether/Hypochlorite 15 min	24,083 ± 0.014	24,106 ± 0.012	4,215 ± 0.010	4,140 ± 0.006
Polyether/Glutaraldehyde 10 min	24,081 ± 0.008	24,079 ± 0.015	4,207 ± 0.015	4,123 ± 0.013
Polyether/Glutaraldehyde 15 min	24,089 ± 0.014	24,097 ± 0.015	4,208 ± 0.015	4,115 ± 0.010
Polysulfide/Control	24,103 ± 0.018	24,092 ± 0.019	4,220 ± 0.029	4,175 ± 0.049
Polysulfide/Hypochlorite 10 min	23,974 ± 0.149	23,998 ± 0.117	4,174 ± 0.016	4,141 ± 0.045
Polysulfide/Hypochlorite 15 min	24,062 ± 0.067	24,067 ± 0.021	4,171 ± 0.041	4,172 ± 0.024
Polysulfide/Glutaraldehyde 10 min	24,102 ± 0.039	24,080 ± 0.018	4,193 ± 0.034	4,154 ± 0.035
Polysulfide/Glutaraldehyde 15 min	23,994 ± 0.044	23,985 ± 0.107	4,188 ± 0.054	4,161 ± 0.031

The Tukey Test was used to analyze the results and showed no significant dimensional changes in polyether and polysulfide impressions in all periods of disinfection with glutaraldehyde and sodium hypochlorite ( $p = 0.05$ ).

Neither polyether nor polysulfide impressions showed any statistically significant differences from their control measurements after soaking in the two disinfectant solutions. The null hypothesis for these experiments was that the mean distances measured in the control group were the same, irrespective of the impression being soaked in either one of the disinfectant solutions. However, after the alginate impressions were disinfected with sodium hypochlorite for 15 minutes a significant change was observed when compared with control group. A reduction in measurement represented alginate shrinkage. Macro and microscopic porosities were also found. The differences attained 0.3 mm ( $3 \times 10^{-4}$  m) mainly in the largest distances (AB and CD).

## DISCUSSION

The effects of different disinfectant solutions and times on three impression materials have been evaluated. There have been disagreements regarding their dimensional stability after the process (Drenon et al.<sup>7</sup>, 1989; Johnson et al.<sup>9</sup>, 1998; Setcos et al.<sup>20</sup>, 1985; Thouati et al.<sup>22</sup>, 1997). According to the specifications provided by the United States Disease Control Centers, chemical disinfectants such as chlorine compounds, formaldehyde, glutaraldehydes, phenols, and iodophors have the potential to eliminate hepatitis, herpes, and HIV viruses in 10 to 30 minutes (Matyas et al.<sup>14</sup>, 1990). In this study, the choice was to evaluate polyether, polysulfide and irreversible hydrocolloid impressions because they are hygroscopic and tend to shrink. Polyether and polyvinyl siloxanes. Sodium hypochlorite and glutaraldehyde were chosen as disinfectants because they are more widely used. Various types of tests were used to evaluate all arch casts, cavities for impressions. In this study, the American Dental Association standardization nº 19 was chosen for standardizing disinfection and impression procedures, because its usefulness in dentistry has been professionally recognized. Individual acrylic resin trays were not manufactured for the impressions because of the risk of water absorption and introduction of other variations. Stainless steel trays were used instead. For the same reason, the gypsum material was not poured.

The problem of disinfecting dental impressions (Lepe et al.<sup>12</sup>, 2002), particularly irreversible hydrocolloid and hydrophilic ones, such as polyethers is a major concern. Herrera et al.<sup>7</sup> (1986); Merchant et al.<sup>15</sup> (1984), Langerwalter et al.<sup>11</sup> (1990) and Matyas et al.<sup>14</sup> (1990) did not find dimensional change after the use of a 0.5% sodium hypochlorite solution for 30 minutes. In this study, the concentration of sodium hypochlorite was 2 times greater, which could explain the differences in the results. However, Minagi et al.<sup>16</sup> (1987) showed that immersion in 2% glutaraldehyde for 60 minutes for irreversible hydrocolloid impression materials did not jeopardize surface details. When it was immersed in 1% sodium hypochlorite for 15 minutes, however, it was possible to observe statistically significant differences in surface alterations, like little cracks and imperfections of dentures. These changes would be transferred to the finished cast and subsequently to the finished denture.

Although no significant linear dimensional changes were observed in polyether impressions for 10 and 15 minutes, previous studies have shown that immersion of this material adversely affects surface details of casts. The same authors recommend that polyethers should not be disinfected with sodium hypochlorite, because they may lose surface details for periods exceeding 5 hours (Bergman et al.<sup>1</sup>, 1989; Johnson et al.<sup>9</sup>, 1998; Drenon et al.<sup>7</sup>, 1989; Johnson et al.<sup>10</sup>, 1998; Setcos et al.<sup>20</sup>, 1985; Owen et al.<sup>17</sup>, 1993; Sawyer et al.<sup>18</sup>, 1974). Disinfection of the polyether impressions using an alcoholic glutaraldehyde solution (2%) and sodium hypochlorite solution (1%), for periods of 10 and 15 minutes led to non-significant variations in all measured distances (AB, CD, AC, BD) without loss of accuracy or surface detail, a result consistent with previous studies. (Johnson et al.<sup>9</sup>, 1998; Drenon et al.<sup>7</sup>, 1989; Johnson et al.<sup>10</sup>, 1998). The results obtained with polysulfide impression disinfections shows no difference in mean values after all times of disinfection with both disinfectant solutions.

The results of this research are important to the dentist to select the appropriate disinfectant solution for specific clinical conditions such as fabrication of study models, removable partial dentures or fixed partial dentures.

## CONCLUSION

Within the limits of this study it can be concluded that disinfect immersion practices

did not influence the dimensional stability of impressions obtained, except when sodium hypochlorite and a 15-minute immersion time were used for disinfecting irreversible hydrocolloid impressions.

#### REFERENCES

1. Bayindir F, Yanikoglu N, Duymus Z. Thermal and pH changes, and dimensional stability in irreversible hydrocolloid impression material during setting. *Dent Mater J*. 2002;21(2):200-9.
2. Bergman M, Olsson S, Bergman B. Elastomeric impression materials. Dimensional stability and surface detail sharpness following treatment with disinfectant solutions. *Swed Dent J*. 1980;4:161-7
3. Bond WW, Favero MS, Petersen NJ, Ebert JW. Inactivation of Hepatitis B virus by intermediate-to-high-level disinfectant chemicals. *J Clin Microbiol*. 1983;18:535.
4. Chong MP, Docking AR. Some setting characteristics of elastomeric impressions materials. *Aust Dent J*. 1969;14:295-301.
5. American Dental Association. Council of Dental Materials and Devices: specification n° 19 for non-aqueous elastomeric dental impression materials. *J Am Dent Assoc*. 1977;94:733-41
6. Dellinger EL, Williams KJ, Setcos JC. Influence of immersion and spray disinfectants on alginate impressions. *J Dent*. 1990;69:364(Abst 2045).
7. Drennon DG, Johnson GH, Powell GL. The accuracy and efficacy of disinfection by spray atomization on elastomeric impressions. *J Prosthet Dent*. 1989;62:468-75
8. Durr DP, Novak EV. Dimensional stability of alginate impressions immersed in disinfectant solutions. *J Dent Child*. 1987;54:45-8.
9. Johnson GH, Chellis KD, Gordon C. Dimensional stability and detail reproduction of irreversible hydrocolloid and elastomeric impressions disinfected by immersion. *J Prosthet Dent*. 1998;79:446-53.
10. Johnson GH, Drennon DG, Powell GL. Dimensional stability of elastomeric impression materials immersed in disinfectant solutions. *J Am Dent Assoc*. 1998;129:1063-7.
11. Langenwaller EM, Aquino R. Dimensional stability of elastomeric impression materials following disinfection with sodium hypochlorite. *J Prosthet Dent*. 2002;88(3):268-76
12. Lepe X, Johnson GH, Berg JC, Aw TC, Stroh GS. Wettability, imbibition, and mass change of disinfected low-viscosity impression materials. *J Prosthet Dent*. 2002;88(3):268-76
13. Look JO. Preliminary results from disinfection of irreversible hydrocolloid impressions. *J Prosthet Dent*. 1990;63:701-7.
14. Matyas, J, Dao N, Caputo AA, Lucatorto FM. Effects of disinfectants on dimensional accuracy of impression materials. *J Prosthet Dent*. 1990;64:25-31.
15. Merchant, V.A, Radcliffe, RM. Preliminary investigation of a method for disinfection of dental impressions. *J Prosthet Dent*. 1984;52:877-9.
16. Minagi, S., Yano, N., Yoshida, K., Tsuru, HSE. Prevention of acquired immunodeficiency syndrome and hepatitis B. II: Gamma radiation method for hydrophilic impression materials. *J Prosthet Dent*. 1987;58:462-5.
17. Owen CP, Gooch B. Dimensional stability of impression materials to disinfection: a review and comparison. *J Dent*. 1993;6:480-94.
18. Rios MF, de Souza R, de Souza S, Rose L. Effects of chemical disinfection on the stability and detail reproduction of elastomeric impression complex. *J Prosthet Dent*. 2002;88(3):268-76.
19. Sato T, et al. Accuracy of cast impressions of three classes of elastomeric impression materials. *J Am Dent Assoc*. 1974;89:1063-7.
20. Schmitt R, Palenik CJ, Hinoura K. Dimensional stability of a polyether dental impression material. *J Dent Res*. 1985;62:244.
21. Cheng M, Palenik CJ, Blumenshine R. Effect of disinfectants on a polysulfide impression material [abstract]. *J Dent Res*. 1986;65:1063-7.
22. Gatti A, Deveaux E, Iost A, Behin P. Dimensional stability of seven elastomeric impressions materials immersed in disinfectants. *J Prosthet Dent*. 1996;76:8-14.
23. Tullner JB, Commette JA, Moon PC. Linear dimensional changes in dental impressions after immersion in disinfectant solutions. *J Prosthet Dent*. 1988;60:725-8.

Recebido para publicação em: 04/11/2005; aceito em: 06/03/2006.

**Endereço para correspondência:**  
LEONARDO HENRIQUE VADENAL PANZA  
Rua Corvina, 22 - Aquários  
CEP 13280-000, Vinhedo, SP, Brasil