Assessment of antimicrobial activity of sodium hypochlorite, calcium hypochlorite and grape seed extract against Enterococcus faecalis

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ABSTRACT

OBJECTIVE: This study aimed to assess the antimicrobial action of calcium hypochlorite [Ca(OCl)2] at concentrations of 2.5% and 6%, and of grape seed extract (GSE) at concentrations of 10%, 30%, and 50%, against Enterococcus faecalis, comparing it to the action of 6% sodium hypochlorite (NaOCl).

METHODS: Saline solution was used as negative control. The inhibition halos of microbial growth were verified by the agar disk diffusion method. Twelve Petri plates were used for seeding with culture medium of approximately 5 mm in thickness. In each plate, 5 disks of pure and sterile antibiogram, soaked in the substances to be tested, were used and taken to the plate containing the seeded bacterial strain. The plates remained in aerobic bacteriological incubator for 24 h at 37°C temperature. After 24 h, the inhibition halos were measured with a digital caliper. Analysis of Variance (ANOVA) was used for statistical analysis followed by Tukey's complementary test, at 5% significance.

RESULTS: The 6% Ca(ClO)2 presented inhibition halo statistically higher than the other solutions (p<0.05), followed by 2.5% Ca(ClO)2, which was statistically similar to 6% NaOCl (p>0.05). The GSE concentrations resulted in lower inhibition halos of active substances and the different concentrations were similar to each other. Lastly, saline solution presented the same inhibition halos in all groups (p<0.05).

CONCLUSION: It may be concluded that 6% Ca(OCl)2 presented higher antimicrobial activity than 6% NaOCl. On the other hand, all GSE concentrations were lower than NaOCl and Ca(OCl)2.

Keywords: calcium hypochlorite; sodium hypochlorite; grape seed extract; Enterococcus faecalis.
INTRODUCTION

Enterococcus faecalis (E. faecalis) is a strong bacterium with ability to survive in root canals as a single organism. It can invade dentinal tubules and resist the action of antimicrobial substances. Because of these features, it is the main microorganism related to endodontic failure [1, 2]. In order to eliminate microorganisms, the cleaning and disinfection of root canals depend on the both action of endodontic instruments and the chemical action of auxiliary substances. The use of endodontic instruments considerably reduces the number of microorganisms within the root canal; however, it is not enough to provide adequate disinfection, so requiring the use of auxiliary chemical substances [3].

The most used solution in Endodontics is sodium hypochlorite (NaOCl), which main component is chloride. In contact with dentin, the NaOCl is dissociated into sodium hydroxide and hypochlorous acid, which are responsible for the ability of tissue dissolution and antimicrobial activity of this solution, respectively [4]. However, it presents cytotoxicity and intense irritation when extrusion occurs through the apical foramen [4, 5]. Moreover, it promotes dentin collagen degeneration [6, 7] and, consequently, negatively interferes with the adhesion of restorative materials to dentin [8, 9].

Considering the disadvantages of NaOCl, it is important to study new auxiliary chemical solutions that may be used during root canal preparation. Calcium Hypochlorite [Ca(ClO)₂] is a white granulate extensively used in water treatment and as a bleaching agent [10]. It is considered chemically stable and has more chloride available than NaOCl [11]. Moreover, its ability to dissolve organic matter [10] and its antimicrobial activity against E. faecalis [11] have been reported with acceptable cytotoxicity and biocompatibility [13].

Another substance that indicated antimicrobial activity in Endodontics and advances for Restorative Dentistry was Grape Seed Extract (GSE), which is a substance richly composed of PAs (proanthocyanidins) [14] from plants such as stems, fruits, seeds, and flowers [15]. Grape Seed Extract induces crosslinking in dentin collagen [16] and it presents low cytotoxicity [17]. It is believed that GSE presents satisfactory antimicrobial activity, considering that Cecchin et al. [18] observed that it presents good ability to eliminate E. faecalis when used in straight canals with manual instruments.

Considering the above, the present study aimed to assess bacterial growth inhibition caused by different auxiliary chemical solutions (GSE, Ca(ClO)₂, and NaOCl) against E. faecalis through the inhibition halo test. The hypothesis tested was that different auxiliary chemical substances would have the same antimicrobial activity.

MATERIALS AND METHODS

Study design

The present work is an experimental laboratory study proposed to determine the antimicrobial activity of the following auxiliary chemical substances: 10% GSE (Mega-Natural, Madera, CA, USA), 30% GSE, and 50% GSE; 6% NaOCl (Natupharma, Passo Fundo, RS, Brazil); and 2.5% Ca(ClO)₂ (Lírios, São Vicente, SP, Brazil) and 6% Ca(ClO)₂. All substances were tested against E. faecalis (American Type Culture Collection 19433) by delimiting inhibition halos through the agar disk diffusion method. The disks used were white for pure antibiogram (no antibiotics) and sterilized by autoclaving (Laborclin Produtos para Laboratórios Ltda., Pinhais, PR, Brazil).

Preparation of substances

The 6% concentration of NaOCl was prepared by the compounding pharmacy Natupharma, Passo Fundo, RS, Brazil.

The 10%, 30%, and 50% concentrations of GSE (Mega-Natural, Madera, CA, USA) and the 2.5% and 6% concentrations of Ca(ClO)₂ (Lírios, São Vicente, SP, Brazil) were prepared from their respective granules. The raw material of substances was diluted in distilled water, resulting in the desired concentrations at the Chemistry Laboratory of the University of Passo Fundo. Different concentrations were prepared at weight/volume ratio and mixed with a magnetic stirrer for 10 minutes.

Preparation of the Inoculum

The E. faecalis strain (ATCC 19433) was reactivated in Brain Heart Infusion broth (BHI, Difco, Kansas City, MO, USA) and incubated at 37°C for 24 hours. Next, it was transferred to a new tube containing BHI broth and incubated for 24 hours more under the same conditions; medium density was adjusted in a spectrophotometer, equivalent to 1.0 in the McFarland scale (3×10⁸ CFU mL⁻¹). After adjusting bacterial concentration, the inoculum was incubated once again at 37°C for 7 hours, to reach exponential growth.

Plate Seeding

Twelve 90-mm Petri plates were used for seeding (n=10). The culture medium of the plates was Agar Mueller Hinton, with approximately 5 mm in thickness. Five disks were used in each plate.

For assistance, a swab was immersed in the BHI inoculum containing the bacterial strain and pressed on the tube walls to remove the excess. Seeding was performed in four different orientations and around the edge of the plate, in order to ensure that the entire medium was seeded.

Application of Disks

The disks remained in the refrigerator up to 2 h before use. Right after seeding, there was a 5-minute interval for the medium to dry at room temperature and to absorb the inoculum before applying the disks.

Each substance tested remained in a sterile Eppendorf tube until use. After 5 minutes, aided by clinical tweezers for sterile cotton (Colgran, São Caetano do Sul, SP, Brazil), the antibiogram disk was previously soaked in the substance for 10 minutes, removed and waited 30 s to be placed on the plate.
When disk placement was finished, the plate was closed and maintained in aerobic bacteriological incubator for 24 h at 37°C temperature. In the first 15 minutes, the plates remained incubated in reverse so the disks would not move, then they were turned with the lid down to prevent the accumulation of droplets on the lid, interfering with the culture medium.

Microbiological Assessment

After 24 h, the plates were removed from the incubator and the inhibition halos from the substances were measured, aided by a digital caliper (Vonder Paquimetro Eletrônico Digital, Curitiba, PR, Brazil).

All plates were read with the help of a light source. The halos were considered from the point where no bacterial growth was visible to the naked eye.

Statistical analysis

The statistical analysis used was ANOVA followed by Tukey’s complementary test, for group comparison, at 5% significance level. For results analysis, the software Stat Plus Analyst Soft Inc. version 6.0 (Vancouver, BC, Canada) was used.

RESULTS

The statistical analysis showed statistically significant difference among groups (α=0.05). The 6% Ca(ClO)₂ presented inhibition halo statistically higher than the other solutions (p<0.05), followed by 2.5% Ca(ClO)₂, which was statistically similar to 6% NaOCl (p>0.05), as described in Table 1.

DISCUSSION

The methodology applied in this research adopted the analysis of the size of bacterial growth display areas, in agar diffusion. It is possible to verify that the 6% Ca(ClO)₂ solution was the concentration that presented the highest inhibition halo. On the other hand, 2.5% Ca(ClO)₂ and 6% NaOCl presented lower halos, but similar to each other. These results show that 2.5% Ca(ClO)₂ presents adequate capacity of bacterial inhibition when compared to high concentrations of NaOCl, considering that some studies prove the efficiency of NaOCl at different concentrations [4, 12]. From these results, the hypothesis under analysis was rejected. There are few studies on Ca(ClO)₂ in the field of Endodontics. This substance was introduced for favoring organic matter degradation at the same proportion of NaOCl and for being more stable [12]. The Ca(ClO)₂ is an alkaline solution with higher chloride content available than NaOCl [11], produced by the dissolution of chlorine gas (Cl₂) in a solution of calcium oxide (CaO) and sodium hydroxide (NaOH), which when diluted in water turns into hypochlorous acid [10, 19]. Its antimicrobial activity may be justified by the high amount of chloride available. High doses of hypochlorous acid are released and invade the bacterial cell wall, destroying the microorganisms present in infections [12]. Using an antimicrobial assessment method different from this study, De-Almeida et al. [12] and Dumani et al. [20] assessed the antimicrobial capacity of Ca(ClO)₂ and proved its effectiveness, considering the solution was able to reduce CFUs without statistical differences with the group using NaOCl.

The antimicrobial efficacy of NaOCl has been extensively discussed in the literature [21, 22]. The NaOCl consists of sodium salts of hypochlorous acid and it is a compound that contains chloride, which may be used as disinfectant [23]. The NaOCl solution presents a high and stable pH, slowly releasing chloride [24]. However, its efficacy may be affected by concentration, temperature, and pH [25, 26]. When the pH is low, the free chloride of NaOCl joins the hypochlorous acid (HOCl), which is more active than hypochlorite anion (OCl⁻) [24].

The GSE was the active solution tested that presented the lowest inhibition halo, therefore with lower antimicrobial activity than Hypochlorite solutions. The interest for this substance is based on antibacterial, antifungal, antiviral, antioxidant, anti-inflammatory, and analgesic properties [17, 27]. The structure of GSE presents PAs [14], which improves the mechanical properties of dentin collagen [16] with proved antimicrobial activity [18]. The phenolic contents present in grape seeds are relatively hydrophobic and communicate with the bacterial cell wall, decreasing membrane stability. Up to now, there are few studies on GSE for endodontic use, and because it is a phytotherapeutic substance, it requires additional studies to prove its effectiveness, provided there is no consensus on the proper concentration for the solution, which does not allow objective conclusions on its use. Studies on the antimicrobial activity

### Table 1. Reduction of Enterococcus faecalis by the agar disk diffusion technique through the direct contact method (n=10).

<table>
<thead>
<tr>
<th>Auxiliary Chemical Solutions</th>
<th>Inhibition Halos (Mean±Standard Deviation)</th>
<th>Tukey’s Statistical Test</th>
</tr>
</thead>
<tbody>
<tr>
<td>Saline solution</td>
<td>0.00±0.00</td>
<td>D</td>
</tr>
<tr>
<td>10% GSE</td>
<td>10.40±1.16</td>
<td>C</td>
</tr>
<tr>
<td>30% GSE</td>
<td>11.97±0.48</td>
<td>C</td>
</tr>
<tr>
<td>50% GSE</td>
<td>13.07±0.65</td>
<td>C</td>
</tr>
<tr>
<td>6% NaOCl</td>
<td>18.52±1.37</td>
<td>C</td>
</tr>
<tr>
<td>2.5% Ca(ClO)₂</td>
<td>22.34±4.96</td>
<td>B</td>
</tr>
<tr>
<td>6% Ca(ClO)₂</td>
<td>29.82±8.23</td>
<td>A</td>
</tr>
</tbody>
</table>

Data are presented as mean±standard deviation. The p values are significant using the analysis of variance in the columns. Different letters represent statistically significant differences in the post-hoc procedure (Tukey’s test). GSE, Grape Seed Extract; NaOCl, Sodium Hypochlorite; Ca(ClO)₂, Calcium Hypochlorite.

The 10%, 30%, and 50% concentrations of GSE showed reduced antimicrobial effectiveness when compared to Ca(ClO)₂ and NaOCl. All GSE concentrations resulted in the lowest inhibition halos among the active substances. Lastly, saline solution, considered as control group, presented lower and different results from the other groups (p<0.05).
of GSE in Endodontics are scarce in the literature. Baydar et al. [28] tested GSE at concentrations of 1%, 2.5%, 5%, and 10% for several bacteria, *E. faecalis* among them, by the agar diffusion method, analyzing inhibition halos. The authors concluded that all concentrations were effective for the bacteria tested. In this study, GSE presented the lowest bacterial reduction from the other solutions, even at a higher concentration of 50%. The GSE may be used as an option of irrigating solution in Endodontics, because it does not interfere with dentin collagen, preserving the quality of dentin substrate in order to obtain an adequate root canal sealing at the moment of filling and, later, for composite resin restoration. Moreover, teeth are an interesting option for the preparation of thin roots or incomplete rhizogenesis, due to low cytotoxicity [17] and their positive properties on dentin collagen [16, 29]. It is important to report that, for Ca(ClO)₂ and GSE solutions, the formation of a precipitate was observed, in which the solid portion was deposited at the bottom of the solution flask. For the solution not to lose its concentration, the flask must be agitated for better diluting the solid portion with the liquid one at the moment of use.

The Ca(ClO)₂ presented favorable result regarding its antimicrobial activity. In vivo researches should be performed to verify the effectiveness of Ca(ClO)₂ and GSE as auxiliary chemical solutions for endodontic treatment.

**CONCLUSIONS**

After the results obtained in this study, it may be concluded that the highest antimicrobial activity was presented by 6% Ca(ClO)₂ followed by 2.5% Ca(ClO)₂, which was similar to 6% NaOCl. The different GSE concentrations did not present statistically significant differences and presented lower antimicrobial capacity than the other substances tested.

**REFERENCES**


Antimicrobial activity of endodontic irrigating solutions | Solgo et al.